

RayBio[®] Human/Mouse/Rat Inhibin B (Beta B subunit) Enzyme Immunoassay Kit

Catalog #: EIA-INB, EIAM-INB, EIAR-INB

User Manual
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Caution:
Extraordinarily useful information enclosed



ISO 13485 Certified

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Please read the entire manual carefully before starting your experiment

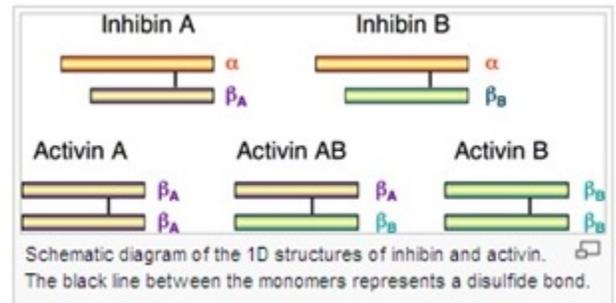
I. Introduction

Inhibin B belongs to TGF-beta superfamily, which members also include Inhibin A, TGF-betas (transforming growth factors), activins, BMPs (bone morphogenetic proteins). Among these factors, inhibins are most closely related to activins.

The activin and inhibin are both dimeric protein complexes in structure, and in each complex, the two monomers are linked to one another by a single disulfide bond. In addition, both complexes are derived from the same family of related genes and proteins but differ in their subunit composition. Inhibins are disulfide-linked heterodimers comprising an alpha-subunit and beta-subunit.

Inhibin B is composed of one alpha subunit and one beta B subunit; Inhibin A is composed of one alpha subunit and one beta A subunit. By contrast, activins are homo- or hetero-dimers, forming activin A (2 beta A subunits), activin B (2 beta B subunits), or activin AB (1 beta A subunit and 1 beta B subunit).

Class	Activity	Complex	Dimer subunits	
			1	2
Inhibin	inhibits FSH secretion	Inhibin A	α	β_A
		Inhibin B	α	β_B
Activin	stimulates FSH secretion	Activin A	β_A	β_A
		Activin AB	β_A	β_B
		Activin B	β_B	β_B



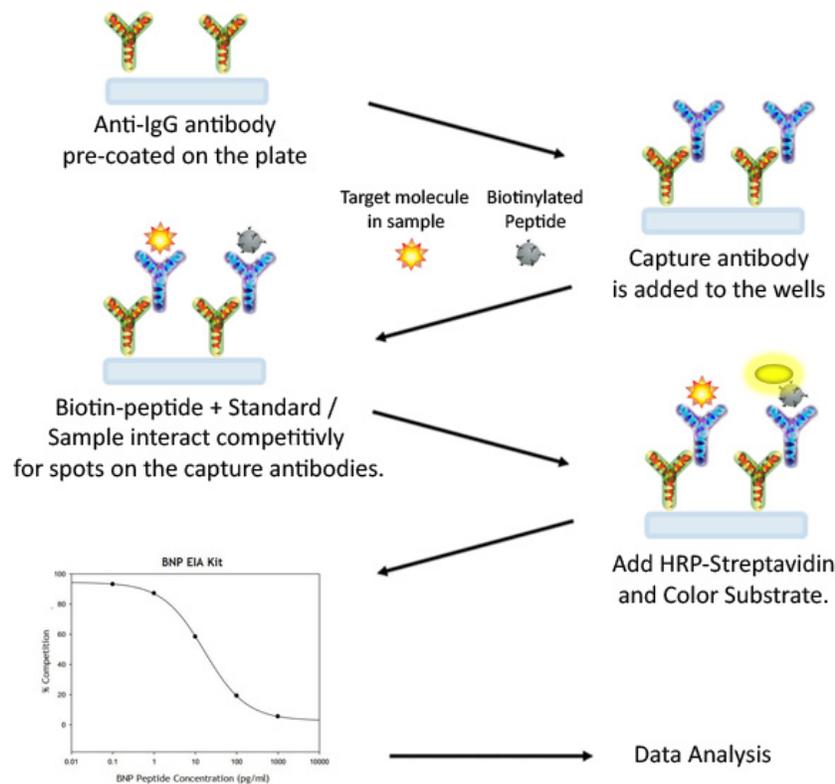
Inhibins have played important roles in regulating reproductive health. Inhibins and activins generally play opposing roles as modulators of the reproductive axis and have multiple roles in the regulation of reproductive physiology in both females and males. Recently the emerging roles for inhibins in tumorigenesis have been reported. Inhibins has been reported as potential tumor suppressors in several different tissues, including ovary, prostate, endometrium and breasts, with loss of inhibin expression or loss of sensitivity linked to tumor initiation and progression as well as poor patient survival.

II. General Description

The RayBio[®] Inhibin B Enzyme Immunoassay (EIA) Kit is an in vitro quantitative assay for detecting Inhibin B peptide based on the competitive enzyme immunoassay principle.

In this assay, a biotinylated Inhibin B peptide is spiked into the samples and standards. The samples and standards are then added to the plate, where the biotinylated Inhibin B peptide competes with endogenous (unlabeled) Inhibin B for binding to the anti-Inhibin B antibody. After a wash step, any bound biotinylated Inhibin B then interacts with horseradish peroxidase (HRP)-streptavidin, which catalyzes a color development reaction. The intensity of the colorimetric signal is directly proportional to the amount of captured biotinylated Inhibin B peptide and inversely proportional to the amount of endogenous Inhibin B in the standard or samples. A standard curve of known concentration of Inhibin B peptide can be established and the concentration of Inhibin B peptide in the samples can be calculated accordingly.

III. How It Works



IV. Storage

The entire kit may be stored at -20°C to -80°C for up to 6 months from the date of shipment. For extended storage, it is recommended to store at -80°C. **Avoid repeated freeze-thaw cycles.** For prepared reagent storage, see table below.

V. Reagents

Component	Size / Description	Storage / Stability After Preparation
EIA Microplate (Item A)	96 wells (12 strips x 8 wells) coated with secondary antibody.	1 month at 4°C*
Wash Buffer Concentrate (20X) (Item B)	25 ml of 20X concentrated solution.	1 month at 4°C
Standard Inhibin B Peptide (Item C)	2 vials of Inhibin B Peptide. 1 vial is enough to run each standard in duplicate.	The first standard: 2-3 days at 4°C Additional dilutions: Do not store
Anti-Inhibin B Polyclonal Antibody (Item N)	2 vials of anti-Inhibin B.	1 month at 4°C
Assay Diluent A (Item D)	30 ml, contains 0.09% sodium azide as preservative. Diluent for standards and serum or plasma.	N/A
Assay Diluent B (Item E)	15 ml of 5X concentrated buffer. Diluent for standards, cell culture media or other sample types, and HRP-Streptavidin.	1 month at 4°C
Biotinylated Inhibin B Peptide (Item F)	2 vials of Biotinylated Inhibin B Peptide, 1 vial is enough to assay the whole plate.	2-3 days at 4°C
HRP-Streptavidin Concentrate (Item G)	600 µl 200X concentrated HRP-conjugated streptavidin.	Do not store and reuse
Positive Control (Item M)	1 vial of Positive Control.	2-3 days at 4°C
TMB One-Step Substrate Reagent (Item H)	12 ml of 3,3',5,5'-tetramethylbenzidine (TMB) in buffer solution.	N/A
Stop Solution (Item I)	8 ml of 0.2 M sulfuric acid.	N/A

*Return unused wells to the pouch containing desiccant pack, reseal along entire edge.

VI. Additional Materials Required

1. Microplate reader capable of measuring absorbance at 450 nm
2. Precision pipettes to deliver 2 μ l to 1 ml volumes
3. Adjustable 1-25 ml pipettes for reagent preparation
4. 100 ml and 1 liter graduated cylinders
5. Absorbent paper
6. Distilled or deionized water
7. SigmaPlot software (or other software which can perform four-parameter logistic regression models)
8. Tubes to prepare standard or sample dilutions
9. Orbital shaker
10. Aluminum foil
11. Plastic wrap

VII. Reagent Preparation

Keep kit reagents on ice during reagent preparation steps.

Note: **Assay Diluent A** should be used for dilution of samples, Item F and Item C when testing **plasma or serum samples**. **1X Assay Diluent B** should be used for dilution of samples, Item F and Item C when testing **cell culture media or other sample types**.

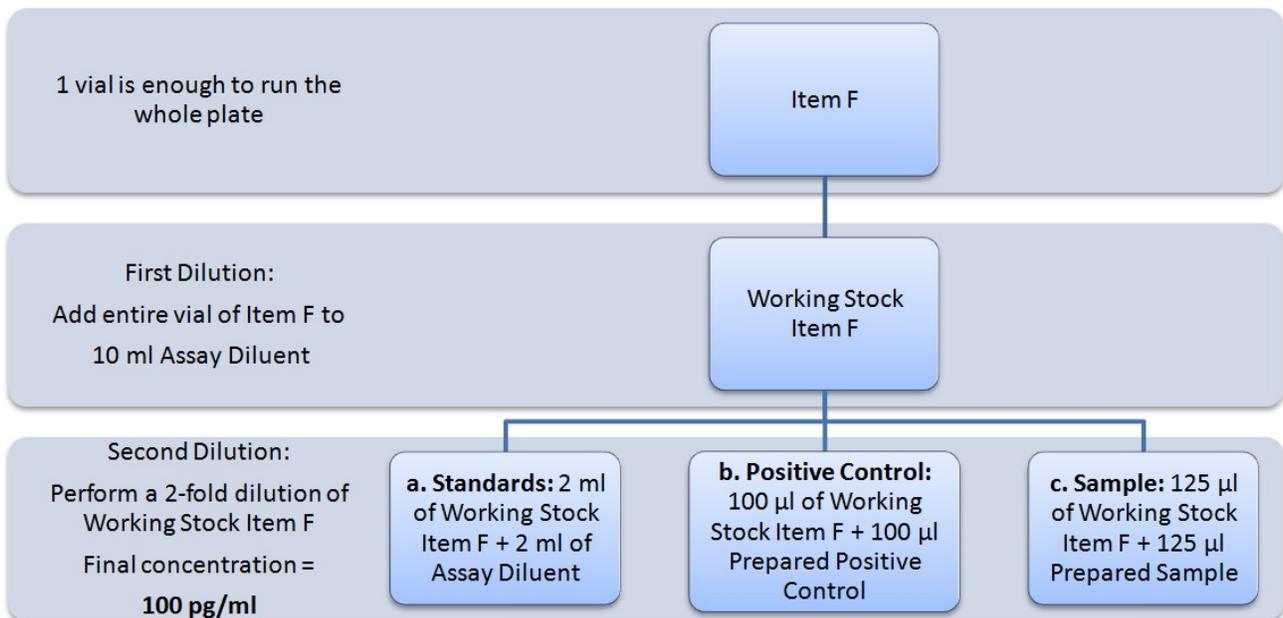
A. Preparation of Plate and Anti-Inhibin B Antibody

1. Equilibrate plate to room temperature before opening the sealed pouch.
2. Label removable 8-well strips as appropriate for your experiment.
3. 5X Assay Diluent B (Item E) should be diluted 5-fold with deionized or distilled water.
4. Briefly centrifuge the anti-Inhibin B antibody vial (Item N) . Then add 50 μ l of 1X Assay Diluent B to the vial to prepare the antibody concentrate. Pipette up and down to mix gently.
5. The antibody concentrate should then be diluted 100-fold with 1X Assay Diluent B. This is your anti-Inhibin B antibody working solution, which will be used in step 2 of Assay Procedure (Section VIII).

Note: The following steps may be done during the antibody incubation procedure (step 2 of Assay Procedure)

B. Preparation of Biotinylated Inhibin B (Item F)

6. Briefly centrifuge the vial of Biotinylated Inhibin B (Item F) before use.
7. See the image below for proper preparation of Item F. Transfer the entire contents of the Item F vial into a tube containing 10 ml of the appropriate Assay Diluent. This is your Working Stock of Item F. Pipette up and down to mix gently. *The final concentration of biotinylated Inhibin B will be **200 pg/ml**.*
 - a. Second Dilution of Item F for Standards: Add 2 ml of Working Stock Item F to 2 ml of the appropriate Assay Diluent. The final concentration of biotinylated Inhibin B will be **100 pg/ml**.
 - b. Second Dilution of Item F for Positive Control: Add 100 μ l of Working Stock Item F to 100 μ l of the prepared Positive Control (Item M). (See section D for Positive Control preparation) The final concentration of biotinylated Inhibin B will be **100 pg/ml**.
 - c. Second Dilution of Item F for samples: Add 125 μ l of Working Stock Item F to 125 μ l of prepared sample (see section E for sample preparation). This is a 2-fold dilution of your sample. The final concentration of biotinylated Inhibin B will be **100 pg/ml**.

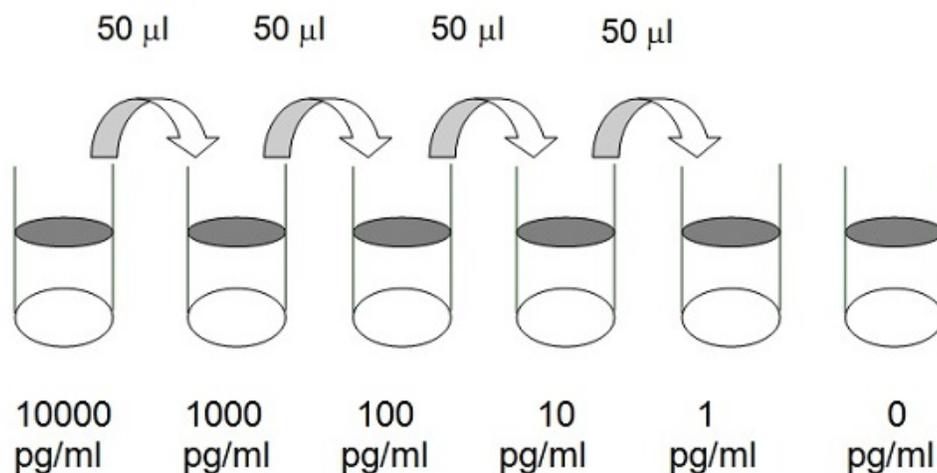


C. Preparation of Standards

- Label 6 microtubes with the following concentrations: 10,000 pg/ml, 1,000 pg/ml, 100 pg/ml, 10 pg/ml, 1 pg/ml and 0 pg/ml. Pipette 450 μ l of biotinylated Inhibin B Item F working solution (prepared in step 7a) into each tube, except the 10,000 pg/ml (leave this one empty).

It is very important to make sure the concentration of biotinylated Inhibin B is 100 pg/ml in all standards.

- Briefly centrifuge the vial of Inhibin B Standard (Item C). Pipette 8 μ l of Item C and 792 μ l of 100 pg/ml biotinylated Inhibin B working solution (prepared in step 7a) into the tube labeled 10,000 pg/ml. Mix thoroughly. This solution serves as the first standard (10,000 pg/ml Inhibin B standard, 100 pg/ml biotinylated Inhibin B).
- To make the 1,000 pg/ml standard, pipette 50 μ l of the 10,000 pg/ml Inhibin B standard into the tube labeled 1,000 pg/ml. Mix thoroughly.
- Repeat this step with each successive concentration, preparing a dilution series as shown in the illustration below. Each time, use 450 μ l of biotinylated Inhibin B and 50 μ l of the prior concentration until the 1 pg/ml is reached. Mix each tube thoroughly before the next transfer.



D. Positive Control Preparation

12. Briefly centrifuge the Positive Control vial (Item M).
13. Refer to step 7b. This is a 2-fold dilution of the Positive Control. The final concentration of biotinylated Inhibin B should still be 100 pg/ml.

The Positive Control is a cell culture media sample that serves as a system control to verify that the kit components are working. The resulting OD will not be used in any calculations; if no positive competition is observed please contact RayBiotech Technical Support. The Positive Control may be diluted further if desired, but be sure the final concentration of biotinylated Inhibin B is 100 pg/ml.

E. Sample Preparation

14. If you wish to perform a 2-fold dilution of your sample, proceed to step 7c. If you wish to perform a higher dilution of your sample, dilute your sample with the appropriate Assay Diluent before performing step 7c.

EXAMPLE (to make a 4-fold dilution of sample):

- a. Dilute sample 2-fold (62.5 μ l of sample + 62.5 μ l of the appropriate Assay Diluent.).
- b. Perform step 7c (125 μ l of working solution Item F + 125 μ l of sample prepared above).

The total volume is 250 μ l, enough for duplicate wells on the microplate. It is very important to make sure the final concentration of the biotinylated Inhibin B is **100 pg/ml**.

Note: Optimal sample dilution factors should be determined empirically, however you may reference below for recommended dilution factors for serum:

Human=2X Mouse=2X Rat=2X.

If you have any questions regarding the recommended dilutions you may contact technical support at 888-494-8555 or techsupport@raybiotech.com.

F. Preparation of Wash Buffer and HRP

15. If Item B (20X Wash Concentrate) contains visible crystals, warm to room temperature and mix gently until dissolved.
16. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1X Wash Buffer.
17. Briefly centrifuge the HRP-Streptavidin vial (Item G) before use.
18. Dilute the HRP-Streptavidin concentrate 200-fold with 1X Assay Diluent B.
*Note: do **not** use Assay Diluent A for HRP-Streptavidin preparation in step 18*

VIII. Assay Procedure

1. Keep kit reagents on ice during reagent preparation steps. It is recommended that all standards and samples be run at least in duplicate.
2. Add 100 μ l of Anti-Inhibin B Antibody (Item N) (See Reagent Preparation step 5) to each well. Incubate for 1.5 hours at room temperature with gentle shaking (1-2 cycle/sec). You may also incubate overnight at 4°C.
3. Discard the solution and wash wells 4 times with 1X Wash Solution Buffer (200-300 μ l each). Washing may be done with a multichannel pipette or an automated plate washer. Complete removal of liquid at each step is essential to good assay performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
4. Add 100 μ l of each standard (see Reagent Preparation Section C), Positive Control (see Reagent Preparation Section D) and sample (see Reagent Preparation Section E) to appropriate wells. Be sure to include a blank well (Assay Diluent only). Cover wells and incubate for 2.5 hours at room temperature with gentle shaking (1-2 cycles/sec) overnight or at 4°C.
5. Discard the solution and wash 4 times as directed in Step 3.

6. Add 100 μ l of prepared HRP-Streptavidin solution (see Reagent Preparation step 18) to each well. Incubate for 45 minutes at room temperature with gentle shaking. It is recommended that incubation time should not be shorter or longer than 45 minutes.
7. Discard the solution and wash 4 times as directed in Step 3.
8. Add 100 μ l of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking (1-2 cycles/sec).
9. Add 50 μ l of Stop Solution (Item I) to each well. Read at 450 nm immediately.

IX. Assay Procedure Summary

1. Prepare all reagents, samples and standards as instructed.
2. Add 100 μ l anti-Inhibin B to each well. Incubate 1.5 hours at room temperature or overnight at 4°C.
3. Add 100 μ l standard or sample to each well. Incubate 2.5 hours at room temperature or overnight at 4°C.
4. Add 100 μ l prepared Streptavidin solution. Incubate 45 minutes at room temperature.
5. Add 100 μ l TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.
6. Add 50 μ l Stop Solution to each well. Read at 450 nm immediately.

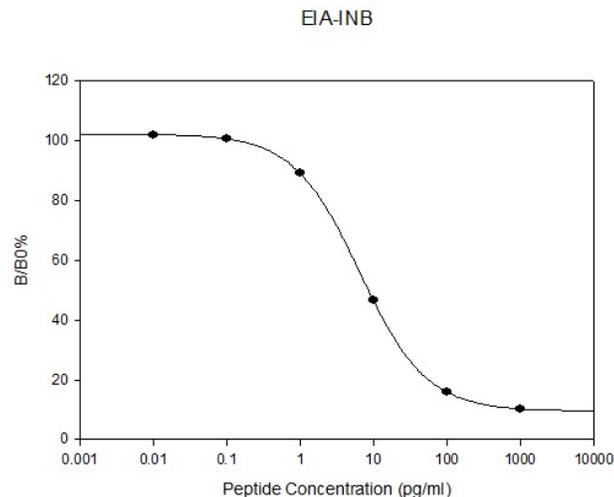
X. Calculation of Results

Calculate the mean absorbance for each set of duplicate stands, controls, and samples and subtract the blank optical density. Plot the standard curve using SigmaPlot software (or other software which can perform four-parameter logistic regression models), with standard concentration on the x-axis and percentage of absorbance (see calculation below) on the y-axis. Draw the best-fit curve through the standard points.

Percentage absorbance = $(B - \text{blank OD}) / (B_0 - \text{blank OD})$ where
B = OD of sample or standard and
B₀ = OD of zero standard (total binding)

A. Typical Data

These standard curves are for demonstration only. A standard curve must be run with each assay.



B. Sensitivity

The minimum detectable concentrations of Inhibin B is 2 pg/ml.

C. Standard Curve Range

1-10,000 pg/ml

D. Reproducibility

Intra-Assay: CV < 10%

Inter-Assay: CV < 15%

E. Assay Diagram

Recommended Plate Layout:

Blank	Blank	SA1	SA1	SA9	SA9	SA17	SA17	SA25	SA25	SA33	SA33
Total Binding	Total Binding	SA2	SA2	SA10	SA10	SA18	SA18	SA26	SA26	SA34	SA34
Standard1	Standard1	SA3	SA3	SA11	SA11	SA19	SA19	SA27	SA27	SA35	SA35
Standard2	Standard2	SA4	SA4	SA12	SA12	SA20	SA20	SA28	SA28	SA36	SA36
Standard3	Standard3	SA5	SA5	SA13	SA13	SA21	SA21	SA29	SA29	SA37	SA37
Standard4	Standard4	SA6	SA6	SA14	SA14	SA22	SA22	SA30	SA30	SA38	SA38
Standard5	Standard5	SA7	SA7	SA15	SA15	SA23	SA23	SA31	SA31	SA39	SA39
Pos Control	Pos Control	SA8	SA8	SA16	SA16	SA24	SA24	SA32	SA32	SA40	SA40

Key:

Blank = Buffer Only

Total Binding = Biotin-Inhibin B only

Standard 1 = 10,000 pg/ml

Standard 2 = 1000 pg/ml

Standard 3 = 100 pg/ml

Standard 4 = 10 pg/ml

Standard 5 = 1 pg/ml

Pos Control = Biotin with Item M

XI. Specificity

This kit targets the beta B subunit, and therefore theoretically detects Activin B and Activin AB in addition to Inhibin B.

Cross Reactivity: This EIA kit shows no cross-reactivity with any of the cytokines tested: Ghrelin, Nesfatin, Angiotensin II, NPY and APC.

XIV. Publications Citing This Product

1. Kyrölähti A, et al. GATA4 regulates Sertoli cell function and fertility in adult male mice. *Molecular and Cellular Endocrinology* Volume 333, Issue 1, 10 February 2011, Pages 85–95
Species: Mouse
Sample Type:
2. Satie AP., et al. Excess Type I Interferon Signaling in the Mouse Seminiferous Tubules Leads to Germ Cell Loss and Sterility. *J Biol Chem.* 2011 Jul 1;286(26):23280-95. doi: 10.1074/jbc.M111.229120
Species: Mouse
Sample Type:

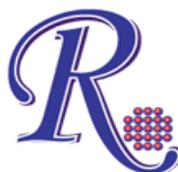
XIII. Troubleshooting Guide

Problem	Cause	Solution
Poor standard curve	<ul style="list-style-type: none"> ○ Inaccurate pipetting ○ Improper standard dilution 	<ul style="list-style-type: none"> ○ Check pipettes ○ Briefly centrifuge Item C and dissolve the powder thoroughly by gently mixing
Low signal	<ul style="list-style-type: none"> ○ Improper preparation of standard and/or biotinylated antibody ○ Too brief incubation times ○ Inadequate reagent volumes or improper dilution 	<ul style="list-style-type: none"> ○ Briefly spin down vials before opening. Dissolve the powder thoroughly. ○ Ensure sufficient incubation time; assay procedure step 2 may be done overnight ○ Check pipettes and ensure correct preparation
Large CV	<ul style="list-style-type: none"> ○ Inaccurate pipetting ○ Air bubbles in wells 	<ul style="list-style-type: none"> ○ Check pipettes ○ Remove bubbles in wells
High background	<ul style="list-style-type: none"> ○ Plate is insufficiently washed ○ Contaminated wash buffer 	<ul style="list-style-type: none"> ○ Review the manual for proper wash. If using a plate washer, ensure that all ports are unobstructed. ○ Make fresh wash buffer
Low sensitivity	<ul style="list-style-type: none"> ○ Improper storage of the ELISA kit ○ Stop solution 	<ul style="list-style-type: none"> ○ Follow storage recommendations in sections IV and V. Keep substrate solution protected from light. ○ Add stop solution to each well before reading plate

RayBio[®] ELISA Kits

Over 3,000 ELISA kits available, visit www.RayBiotech.com/ELISA-Kits.html for details.

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